



## Bluetongue in Denmark 2008

**Rasmussen, Lasse Dam; Rasmussen, Thomas Bruun; Belsham, Graham; Strandbygaard, Bertel; Bøtner, Anette**

*Publication date:*  
2009

*Document Version*  
Publisher's PDF, also known as Version of record

[Link back to DTU Orbit](#)

*Citation (APA):*  
Rasmussen, L. D., Rasmussen, T. B., Belsham, G., Strandbygaard, B., & Bøtner, A. (2009). *Bluetongue in Denmark 2008*. Poster session presented at 3rd Annual Meeting of EPIZONE, Antalya, Turkey.

---

### General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.



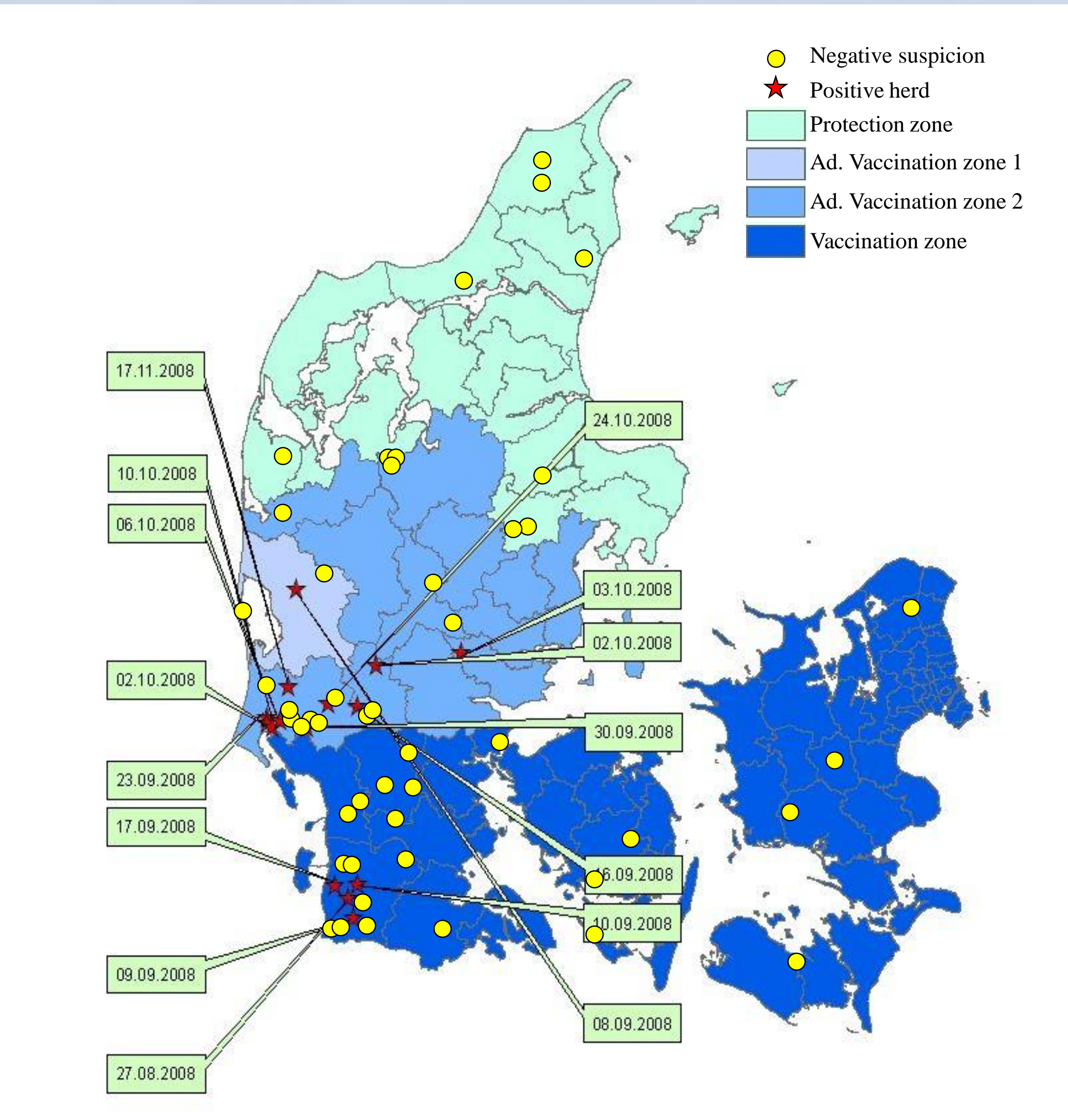


# Bluetongue in Denmark 2008

**Lasse Dam Rasmussen, Thomas Bruun Rasmussen, Graham J. Belsham, Bertel Strandbygaard & Anette Bøtner**

Technical University of Denmark, National Veterinary Institute, Division of Virology, Lindholm DK-4771 Kalvehave, Denmark

## Overview of Bluetongue in Denmark 2008



## Background

The first case ever of bluetongue (BT) in Denmark was recorded in October 2007, so the biting midge season in spring 2008, was awaited with some anxiety, due to the rapid spread of the BTV serotype 8 epidemic in the countries south of Denmark.

## First outbreak in 2008

The first outbreak in Denmark 2008 was detected on August 27 in a cattle herd in Bredebro, which is located in the southern part of Denmark, approximately 20 km north of the German border. Initially two animals were suspected of having BT based on clinical symptoms so EDTA blood and serum samples were submitted to our laboratory. One of the animals tested positive by ELISA (OD% 15.52 1.4) and real time PCR<sup>(1)</sup> (Ct 27.1 0.4)

Species		Antibodies ELISA		Real time RT-PCR	
		+/	OD%	Pool #	Individual
cattle	-				
cattle	+	15,496		1	
cattle	?	43,182		26,75	
cattle	+	3,375			24,53
cattle	+	39,738			
cattle	+	31,887			
cattle	-			2	
cattle	+	25,069			
cattle	+	39,187			
cattle	-				
cattle	+	1,928			28,52
cattle	?	46,006		3	
cattle	-			30,88	
cattle	-				
cattle	-				
cattle	-			4	
cattle	-				
cattle	-				
cattle	+	38,636		5	
cattle	-				
cattle	-				
cattle	+	25,138			23,04
cattle	+	7,645			
cattle	-			6	
cattle	-			25,51	
cattle	+	35,124			
cattle	+	29,477			
cattle	+	24,862		7	
cattle	+	30,441			
cattle	+	21,832			
cattle	-			8	
cattle	+	39,463			
cattle	?	42,7			
cattle	?	43,802			
cattle	-			9	
cattle	+	36,295			
cattle	-				
cattle	+	24,931			
cattle	-				
cattle	-			10	
cattle	-				
cattle	+	35,606			
cattle	+	24,656			
cattle	-			11	
cattle	-				
cattle	-				
cattle	-			12	
cattle	-				
cattle	-				
cattle	-				
cattle	-			13	
sheep	+	4,614			
sheep	+	7,851			
sheep	-				
sheep	+	20,868			14
sheep	+	32,094			
sheep	+	8,678			
sheep	+	14,738			
sheep	?	5,372			
sheep	?	45,248			
sheep	+	9,091			
sheep	+	3,237			
sheep	+	13,981			15

To evaluate the extent of this outbreak it was decided to test the remaining animals in pools from 5 samples.

Three pools were found positive for BT virus (BTV) by real time PCR. In each of these pools one sample of the five was positive for BTV RNA.

The animals were also tested individually for the presence of antibodies against BTV by ELISA.

Of the 75 animals tested, 35 were found positive for BTV specific antibodies. Of these 24 were cattle and the rest were sheep. The three PCR positive cows had high levels of anti-BTV antibodies (OD% 4.32 2.97) whereas the 21 antibody positive but PCR negative cows had lower levels of antibody (OD% 33.03 8.49).

The latter results are probably due to the fact that vaccination against BTV took place 9 days prior to the collection of blood and several animals had seroconverted in the intervening days.

In total in Denmark during 2008 some 15 outbreaks of BT were registered. Out of approx. 65 clinical suspicions 11 were found positive and the remaining four outbreaks were discovered by the routine surveillance of bulk milk. All outbreaks were located in the south western part of the country but several outbreaks were north of the original vaccination zone resulting in two extensions of the vaccination zone during fall 2008. All outbreaks in Denmark during 2008 were caused by BTV-8.

From the first outbreak, a newly infected calf (still sero-negative) was brought to our animal facilities at Lindholm in order to be able to follow the development of anti-BTV antibodies and the level of viral RNA in the blood.

Blood was sampled frequently for a period of three months and analyzed by ELISA, real time PCR and virus isolation. The calf seroconverted one week after bluetongue was diagnosed and subsequently the level of antibodies increased for a month. Real time PCR values remained at an almost constant level throughout the entire three month period, Ct value at diagnosis was 22 and at slaughter the Ct was 25. Virus could be cultured in BHK cells until day 17 after diagnosis.

	Days after BTV diagnosis					
	0	7	17	27	34	81
Virus detection <sup>a)</sup>	22	22	23	24	23	25
Antibodies <sup>b)</sup>	neg	12,329	6,517	3,347	3,082	ND
Virus isolation <sup>c)</sup>	+	+	+	-	-	-

a) Ct values of real time RT-PCR performed on RNA purified from EDTA stabilised blood

b) OD% of Elisa test performed on serum

c) Virus were cultured in BHK-21 cells from washed blood

## Vaccination experiment

The clinical symptoms in the first herd were observed in connection with vaccination against BT and the blood, from the two first animals, submitted for examination was sampled approx. 30 min. after vaccination, which raised the question of whether the positive PCR results could be due to the vaccine.

In order to address this issue, a study was performed in which blood samples were collected at short time intervals immediately before vaccination until 96 h after vaccination (sampling times: 0.25, 1, 4, 24, 48 and 96 h).

Detection of Bluetongue virus and antibodies in newly vaccinated calves

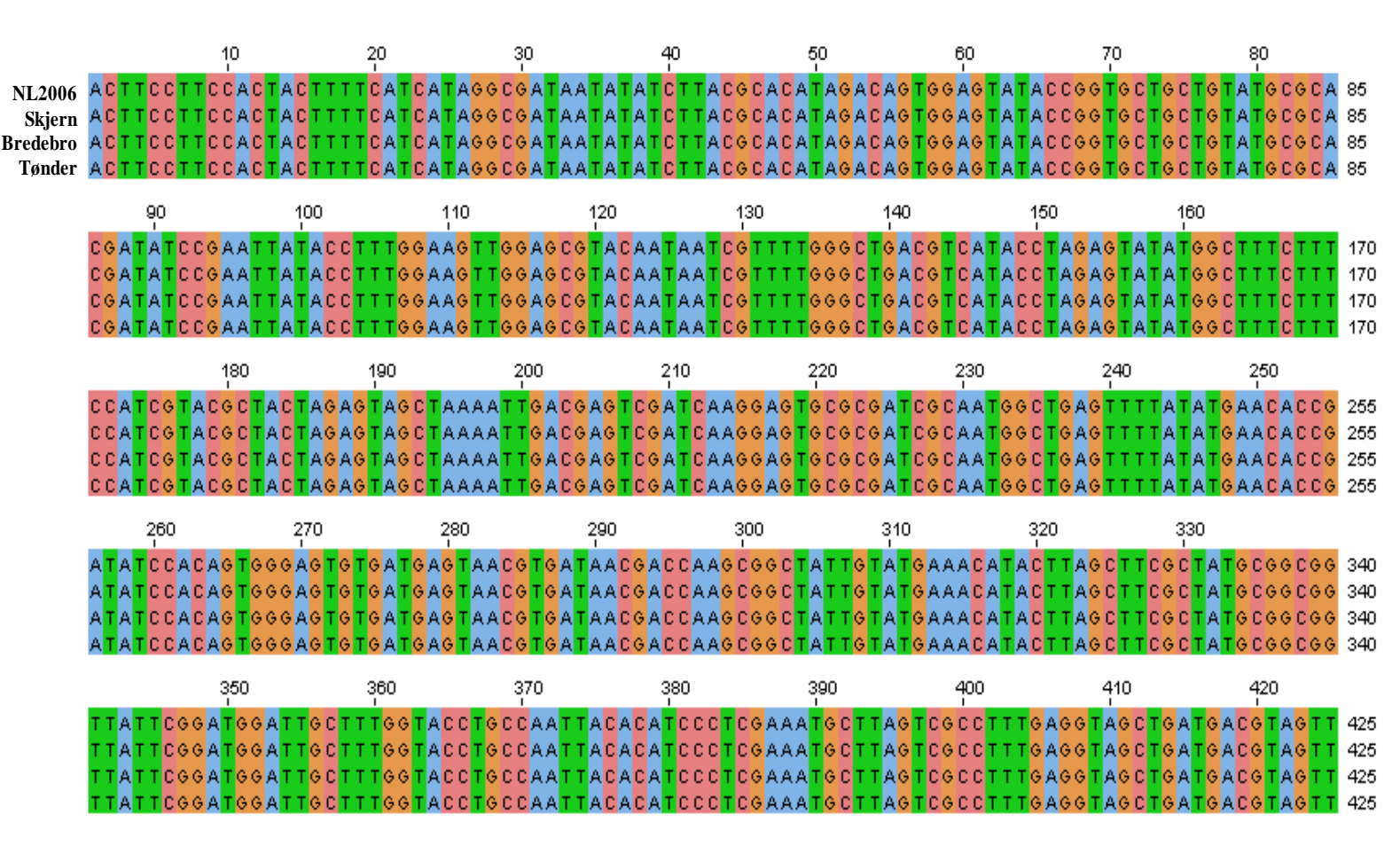
Animal ID	Sampling time after Vaccination (h)						
	0	0,25	1	4	24	48	96
4748							
4858							
4773							
4836							
4729							
4749							
NO Ct							
							ND
Antibodies							
4748							
4858							
4773							
4836							
4729							
4749							
None							

## Serotyping & sequencing

Bluetongue virus from all outbreaks were serotyped by real time PCR (Taqvet BTV8, LSI, Lissieu, France) and found to be BTV 8.

Sequencing of a part of segment 2 (using primer set 8W<sup>(2)</sup>) was performed on BTV RNA from three outbreaks which were believed to be representative of all outbreaks. The most northern (Skjern, 08.09.2008), the most southern (Tønder, 09.09.2008) and the first outbreak (Bredebro, 27.08.2008). See map under the dates for geographic location.

Sequences were identical from all three outbreaks and had 100% homology with the BTV8 isolates circulating in Holland in 2006.



## References

- Shaw, A.E., Monaghan, P., Alpar, H.O., Anthony, S., Darpel, K.E., Batten, C.A., Guercio, A., Alimena, G., Vitale, M., Bankowska, K., Carpenter, S., Jones, H., Oura, C.A., King, D.P., Elliott, H., Mellor, P.S., Mertens, P.P.C., 2007. Development and validation of a real-time RT-PCR assay to detect genome bluetongue virus segment 1. J. Virol. Methods 145, 115–126.
- Mertens, P.P.C., Maan, N.S., Prasad, G., Samuel, A.R., Shaw, A.E., Potgieter, A.C., Anthony, S.J., Maan, S., 2007. The design of primers and use of RT-PCR assays for typing European BTV isolates: differentiation of field and vaccine strains. J. Gen. Virol. 88, 2811–2823.

## Contact Info:

Lasse Dam Rasmussen  
LDRA@vet.dtu.dk